Single Market Programme (SMP Food)

EU co-funded programme for the surveillance of Avian Influenza in poultry and wild birds for 2024

EUROPEAN HEALTH AND DIGITAL EXECUTIVE AGENCY (HADEA)
Department A Health and Food Unit A2 EU4Health/SMP

SUBMISSION FORM: DESCRIPTION OF THE ACTION
(Annex 1 – Description of the action (part B))
Avian Influenza surveillance Programme

Countries seeking an EU financial contribution for the implementation of national programmes for eradication, control and/or surveillance of animal diseases and zoonosis shall submit this Form (*Annex 1 - Description of the action (part B)*) **completely filled in**, **by the 31 May** of the year preceding its implementation (*Part 2.1 of Annex I to the Single Market Programme Regulation*).

Applicant shall provide information on each question contained in the Form. The information filled in the Form, shall be clear, concise, consistent and complete.

For questions on the information requested in this Form, please contact: please contact: HADEA-VET-PROG@ec.europa.eu.

For more information or questions on the eGRANTS Portal Submission System, please access GoFund or contact the IT Helpdesk.

APPLICANT (Name of EU / non-EU country)	
Disease	**AVIAN INFLUENZA**
Species	**Poultry** ☒ **Wild birds** ☒
Implementation Year	**2024**

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Job type within the CA	Head of Epidemiology and Animal Unit

Avian Influenza Programme – 2024

RELEVANCE

1.1 Background and general objectives (*in relation to the Call*)

By submitting this programme, the Member State (MS) attests that the relevant provisions of the EU legislation will be implemented during its entire period of approval, in particular:

- **Commission Delegated Regulation (EU) 2020/689** on 17 December 2019 supplementing Regulation (EU) 2016/429 of the European Parliament and of the Council as regards rules for surveillance, eradication programmes, and disease-free status for certain listed and emerging diseases **(**OJ L 174, 3.6.2020, p. 211–340)

This programme contributes to the maintenance of a high level of health and safety of animals and of humans through early detectiom of avian influenza outbreaks and enforcing subsequent control and eradication measures. ***(maximum 200 characters)***

1.2 Needs and specific objectives

Please give a short description of the programme and in particular how the Objectives for surveillance in poultry and wild birds were met (e.g. please provide a short description of the designed surveillance and indicators to meet each of the objective)

1. Early detection of highly pathogenic avian influenza (HPAI) in poultry.
2. Early detection of HPAI in wild birds providing for:
   (a) an early warning for possible HPAI introduction into poultry, in particular when viruses enter the Union through migratory movements of wild birds;
   (b) information for the assessment of risks for virus spread following findings of HPAI in wild birds.
3. Detection of HPAI in poultry species which generally do not show significant clinical signs.
4. Detection of circulating low pathogenic avian influenza viruses (LPAIV) that may easily spread between poultry flocks in particular in areas with a high density of poultry establishments in view of their potential to mutate to HPAI in order to:
   (a) identify clusters of infection with LPAIV; and
   (b) monitor the risk of spread of LPAIV by movements of poultry and by fomites in certain production systems at risk.
5. Contribution to increased knowledge on HPAI and LPAIV posing a potential zoonotic risk.

The programme is carried out according to annex II to Commission Delegated Regulation (EU) 2020/689. Both passive surveillance in wild birds and active surveillance in poultry will focus on areas at high risk for introduction of avian influenza viruses. In addition, active surveillance in poultry will also be carried out in areas with a high density of poultry production establishments. Early detection of HPAI in poultry will be based on detection of clinical signs of disease, increased mortality rates, decreased feed and water consumptions and decreases in production.

(maximum 500 characters)

1.3 Complementarity with other actions — European added value

Explain how the project builds on the results of past activities carried out in the field.

Illustrate the European dimension of the activities: trans-national dimension of the project; impact/interest for a number of EU countries; possibility to use the results in other countries, potential to develop mutual trust/cross-border cooperation among EU countries, EU and non-EU countries, etc.

Which countries will benefit from the project (directly and indirectly)?

HPAI is a zoonotic transboundary animal disease and currently, since the fall of 2021, there is widespread circulation of virus, mostly subtype H5N1, within the EU territory, having a huge impact in wild birds and poultry populations. This programme, by aiming at early detection of infection, allows for a quick and effective response through the implementation of appropriate biosecurity and outbreak control and eradication measures. Hence, it is beneficial for all Member States, for trade within the Union and with third countries and for animal health and welfare as well as for public health.

(maximum 500 characters)

1.4 Target population and area of the implementation

Describe areas of the implementation of the programme activities (e.g. passive surveillance; active surveillance (clinical examination of herds; sero-surveillance); vaccination (if implemented). If possible, provide maps in the Annex.

If applicable, explain factors/considerations taken into account when deciding on the surveillance type and area of its implementation; in case of vaccination, explain boundaries and size of the vaccination area.

Describe target animals and animal population size both for poultry and wild birds (species, number of holdings or herds or establishment as appropriate and animals) - Fill in **Table 1** (as appropriate) **in the Annex** to this Form.

1.4.1 Passive surveillance in poultry

Avian influenza is a notifiable disease and all increased mortalities and other signs of serious diseases or significantly decreased production rates with an undetermined cause in poultry have to be reported and are duly investigated by The General Directorate for Food and Veterinary (DGAV) in the mainland and Regional Directorates of Agriculture (DRA) in the Autonomous Regions of Açores and Madeira. This early detection system of HPAI is implemented throughout the whole country and includes all poultry categories.

1.4.2 Active surveillance in poultry

Direct and indirect contacts between wild birds and poultry are the main factors associated to introduction of avian influenza virus in poultry establishments. Please see map of higher risk areas (HRA) for introduction of avian influenza virus in Portugal in the annex. This map includes only the mainland for the autonomous regions are remote areas in the Atlantic Ocean and not usually included in Eurasian wild bird migration routes associated to avian influenza

virus introduction into the European Union Territory. Besides this risk factor, areas with high density of poultry establishments, mostly located in Centro and Lisboa e Vale do Tejo regions, will also be targeted due to greater risk regarding the dissemination of infection in the event of an avian influenza outbreak. Sampling will be focused on the most frequently affected as well as the longer-lived and larger production cycle poultry categories, namely ducks, laying hens (including free range), gallinaceous and waterfowl game birds, quails and turkeys which are at higher risk of infection.

Risk-based complementary surveillance for HPAI in poultry species which generally do not show significant clinical signs as well as risk- based surveillance to identify clusters of Low Pathogenic Avian Influenza (LPAI) infected establishments will be implemented according to sections 5 and 6 of Annex II of Commission Delegated Regulation (EU) 2020/689.

Under this programme, poultry species and categories included in section 7 Annex II of Commission Delegated Regulation (EU) 2020/689 will be sampled: breeding ducks, fattening ducks, game birds waterfowl, quails, laying hens (including free range), fattening turkeys and game birds gallinaceous.

Risk based sampling will be carried out focusing on establishments located in the municipalities and parishes located in higher risk areas for the introduction of avian influenza virus and/or in the municipalities at higher risk for disease dissemination, that is, those with higher density of poultry establishments.

a) Risk-based complementary surveillance for Highly Pathogenic Avian Influenza in poultry species which generally do not show significant clinical signs, according to point 2 of section 7 of Annex II of Commission Delegated Regulation (EU) 2020/689:
   All poultry establishments, keeping target poultry species and categories, located in HRA for the introduction of avian influenza virus will be sampled. For poultry establishments, not located in HRA, keeping duck breeders, fattening ducks, game birds waterfowl and quails, the number of establishments to be sampled was calculated considering an estimated prevalence of 5% and 99% confidence level.
b) Risk- based surveillance to identify clusters of Low Pathogenic Avian Influenza infected establishments:
   Sampling will be carried out according to point 3 of section 7 of Annex II of Commission Delegated Regulation (EU) 2020/689. All establishments, keeping target poultry species and categories, located in HRA for the introduction of avian influenza virus will be sampled. In the remaining territory, sampling will be focused in areas of higher poultry density and an estimated prevalence of 5% and 95% confidence level was considered.

For poultry establishments data please see Annex 1.

1.4.3 Passive surveillance in wild birds

Wild birds, in particular migratory water birds, that have been shown to be at a higher risk of becoming infected with and transmitting the HPAI virus, the so called "target species" according to EFSA's Scientific Opinion (2017) will be specifically targeted. Areas close to the sea, lakes and waterways, especially when close to zones of high density of poultry holdings, will be targeted. Additionally, a small number of samples was foreseen for the autonomous regions of Açores and Madeira to allow some degree of monitoring regarding the possibility

of introduction of HPAI virus through other migratory pathways, namely from North America or Africa, which may reach these non-contiguous territories.

Samples will also be taken wherever and whenever significant increases of morbidity and mortality in wild birds occur. Sampling will be carried out in injured, diseased, or dead birds. Additional investigations may be carried out in wild birds in the areas in case of detection of HPAI outbreaks in poultry and/or wild birds or in neighbouring areas.

Data regarding wild birds populations in Portugal is available at https://environment.ec.europa.eu/topics/nature-and-biodiversity/birds-directive_en#reporting. In this webpage please see Reporting period 2013-2018 – National summaries (PDF). Alternatively, please see annex V – Wild birds population data.

1.5 Epidemiological situation

Describe the current epidemiological situation, mention risks/factors which may contribute disease introduction and spread; indicate likelihood for disease introduction and spread from the neighbouring countries.

Provide results of disease surveillance over the last five years for both poultry and wild birds - Fill in **Table 2** (as appropriate) **in the Annex** to this Form.

The main risk factor for disease introduction is contact, direct or indirect, with infected wild birds, namely migratory aquatic and marine wild birds. According to EFSA's Scientific Opinion on Avian Influenza (EFSA Journal 2017;15(10):4991), the wild bird migratory routes mostly associated to the risk of introduction of HPAI virus into the European Union territory are North-eastern and Eastern routes and Portugal is geographically located outside the usual pathways of these routes. However, being part of the southern and north-western flyways, HPAI virus introduction by wild birds originating from Africa, eastern Mediterranean or northern Europe is possible.

On the other hand, subsequently to AI virus introduction, there is an increased risk of disease dissemination in poultry high density areas, especially when biosecurity levels are low, or biosecurity procedures are not strictly followed. This is closely related to the poultry production sector structure which is mainly organized in integrated companies.

When carrying out risk assessment, besides results obtained by the surveillance programme, epidemiological situation in other Member States, especially Spain, and data from the Wild Birds Migration Tool and EFSA's Bird Flu Radar are also considered.

During the last five years, namely in 2021 and 2022, 32 outbreaks of H5N1 HPAI were detected in Portugal: 13 in wild birds and 19 in kept birds, including poultry and captive birds. Except for one outbreak in a fattening ducks establishment sampled by active surveillance, all outbreaks were detected by passive surveillance.

2. QUALITY

2.1 Concept and methodology (Programme activities/measures)

The programme activities/measures shall be clear, suitable to address the needs and to achieve desired outcomes / impact. They must be adapted to disease situation/risk and feasible in terms of the capacities for their implementation.

Clearly describe planning and implementation arrangements/methodology; ensure technical quality and logical links between identified problems/needs and solutions/activities proposed to help improvement; mention timeline for the implementation of specific activities. Further instructions are provided below.

2.1.1 Disease surveillance

Describe disease surveillance (e.g. active (clinical examination of herds; sero-surveillance); passive). For each type of surveillance to be implemented describe: calculations of targets (per risk area if applicable), criteria to include a holding (or herd) and an animal in active surveillance; how holdings will be selected; frequency and timeline of the implementation of clinical examinations (including interval between visits); sampling scheme / sampling strategy, type of samples, who performs clinical examination and sampling; documented procedures for clinical examination, sampling, collection and delivery of samples.

Describe case definition.

a) Passive surveillance in poultry

The aim of passive surveillance is to identify clinical suspicions of avian influenza virus infection in all poultry species and/or production categories. This suspicion may arise from the observation of a sudden increase in flock mortality, clinical signs suggestive of disease or changes in production parameters and feed and/or water consumption. About increased mortality and changes in production parameters, there are no absolute values to be considered; what matters is the detection of changes in the characteristic patterns of each flock.

All suspicions of AI virus infection must be investigated by an official veterinarian who will visit the holding concerned for clinical examination of the birds, examination of the flock register(s) and sampling.

Following that visit, if the official veterinarian considers that the clinical picture and/or the production changes in the flock support a suspicion, in addition to taking samples for laboratory tests he/she shall place the holding under official surveillance as provided for in the operations manual of the avian influenza contingency plan for suspicious situations.

The following samples shall be taken:

(a) 5 recently dead birds

(b) 20 tracheal/oropharyngeal swabs + 20 cloacal swabs from sick birds.

If no mortality occurs, only those samples referred to under (b) must be taken.

b) Active surveillance in poultry

Under this programme, poultry species and categories included in section 7 Annex II of Commission Delegated Regulation (EU) 2020/689 will be sampled: breeding ducks,

fattening ducks, game birds waterfowl, quails, laying hens (including free range), fattening turkeys and game birds gallinaceous.

Risk based sampling will be carried out focusing on establishments located in the municipalities and parishes located in higher risk areas for the introduction of avian influenza virus and/or in the municipalities at higher risk for disease dissemination, that is, those with higher density of poultry establishments. All poultry establishments, keeping target poultry species and categories, located in higher risk areas (HRA)for the introduction of avian influenza virus will be sampled. For poultry establishments, not located in HRA, keeping duck breeders, fattening ducks, game birds waterfowl and quails, the number of establishments to be sampled was calculated considering an estimated prevalence of 5% and 99% confidence level. For poultry establishments, not located in HRA, keeping laying hens, free range laying hens, fattening turkeys and gallinaceous game birds, an estimated prevalence of 5% and 95% confidence level was considered.

Number of sampling events by poultry species/category:

Duck breeders, waterfowl game birds, laying hens and free range laying hens: twice/year;

Fattening ducks: 2 or 3 times per year;

Fattening turkeys, quails, gallinaceous game birds: once/year.

Sample type:

Laying hens, including free range, and fattening turkeys: blood

Duck breeders, waterfowl game birds, gallinaceous game birds and quails: oropharyngeal or cloacal swabs.

Number of establishments to be sampled:

a) Duck breeders – 2 establishments located at higher risk areas,
b) Fattening ducks – 20. Of which 6 are located at higher risk areas,
c) Game birds waterfowl – 2, one is located at a higher risk area,
d) Quails – 18, all not located in higher risk areas,
e) Laying hens – 58. Of which 5 are located at higher risk areas,
f) Free range laying hens – 40. Of which 2 are located at higher risk areas,
g) Fattening turkeys – 45. Of which 14 are located at higher risk areas,
h) Farmed game gallinaceous – 32. Of which 8 are located at higher risk areas.

Sampling is carried out by official veterinarians, or by other DGAV services staff under supervision of official veterinarians, either at poultry establishments or at slaughterhouses, and DGAV personnel is responsible for samples delivery to the laboratory.

Case definition is according to article 9 and annex I to Commission Delegated Regulation nº 2020/689.

c) Passive surveillance in wild birds

Surveillance design is according to sections 4 and 8 of annex II to Commission Delegated Regulation nº 2020/689 and is described in point 1.4.3. In this case, samples to be collected are wild birds' cadavers or oropharyngeal swabs and testing will be carried out by real-time RT-PCR. Samples may be tested in pools of up to 5 if collected at the same sampling event on

wild birds of the same species and origin. A total of 220 samples are to be collected and the number of samples by region is shown on table 1c.

Sampling is carried out by official veterinarians, or by other DGAV services staff under supervision of official veterinarians or by field workers of the Institute for Nature Conservation and Forestry, IP (ICNF), by special brigades of the National Republican Guard and Police Force and by the municipality veterinarians. Nature conservation organizations, bird ringing teams, hunters and ornithologists may also collaborate in the sample collection.

2.1.2 Laboratory testing

Describe tests and testing schemes/hierarchy used; in particular explain the testing scheme.

Mention testing laboratories and tests they perform.
Describe assurance of the quality of the results produced by these laboratories (it is sufficient to indicate laboratory quality assurance schemes in place).

Fill in **Table 1** (as appropriate) **in the Annex** to this Form.

All laboratory testing is done at the National Institute of Agricultural and Veterinary Research, IP (INIAV), which is the National Reference Laboratory for Animal Health. The testing scheme is the following:

a) Passive surveillance samples, in kept and wild birds, are tested for the presence of AI virus by real-time RT-PCR. If positive, subtyping and pathotyping by sequencing or real-time RT-PCR are then carried out.
b) Active surveillance samples: sera obtained blood samples are tested by ELISA and if positive, HI assays are carried. If HI is positive, an official veterinarian shall visit the sampled establishment to carry out clinical inspection of the flock and collect samples for testing by real-time RT-PCR. If these samples are positive, subtyping and pathotyping by sequencing or real-time RT-PCR are then carried out. Oropharyngeal and cloacal swabs are tested as described for passive surveillance samples.

INIAV regularly participates, with favourable results, in laboratory proficiency schemes organized by the European Reference Laboratory for Avian Influenza and Newcastle Disease, including serology assays, real-time RT-PCR and sequencing.

ELISA assays are accredited according to EN 17025.

2.1.3 Measures in case of disease suspicion and confirmation

Describe measures to be implemented in case of disease suspicion and confirmation (detailed references to the provisions of relevant Union legislation to be implemented in case of disease suspicion and confirmation are sufficient).

Avian influenza is a notifiable disease and measures to be taken in the event of a suspicion or confirmation of an outbreak of HPAI are those foreseen in Part III, Title II, chapter 1 of that regulation as well as in Part II, Chapter 1 of Delegated Regulation (EU) 2020/687.

a) **Poultry**: any poultry establishment suspected to be infected with a HPAI virus shall be placed under official surveillance and preliminary movement restriction of birds and products is enforced. Additional biosecurity measures will also be implemented. DSAVR/DRA will carry out an investigation, including clinical examination and sampling as well as an initial epidemiological inquiry. In case of confirmation of infection with HPAI virus, control measures foreseen in articles 12 to 20 of Delegated Regulation (EU) 2020/687 will be implemented. Furthermore, protection and surveillance zones will be established around the infected establishment and surveillance according to article 21 of the aforementioned regulation. Within these restriction zones, an inventory of all establishments keeping poultry, including the species, categories, and estimated number of animals in each establishment, will be carried out and movements of birds and products shall be restricted. Also, additional surveillance and increased biosecurity measures will also be implemented. Within the protection zone, the implementation of measures will be according to articles 25 to 27 and 38 to 39 of Delegated Regulation (EU) 2020/687. Within the surveillance zone, measures will be implemented according to articles 40 to 42, 54 and 55 of the previous mentioned delegated regulation.

Repopulation of the infected poultry establishments will be carried out according to articles 57 to 61 of Delegated Regulation (EU) 2020/687.

In case of detection of LPAI infected establishments, measures foreseen in Decree-Law nº 39:209 of May, 14th, 1953 will be implemented. Besides placing infected establishments under official control, these may include carrying out an epidemiological inquiry, killing and destruction of infected birds, destruction of infected products and materials, cleaning and disinfection procedures under official supervision and animal and products movement restrictions.

b) **Wild birds**: in the event of a suspicion or confirmation of an outbreak of HPAI in wild birds measures will be taken according to article 70 of Regulation (EU) 2016/429 and articles 62 to 67 of Delegated Regulation (EU) 2020/687.

2.1.4 Data collection, management and analysis

Describe surveillance data collection, management and analysis, including spatial analysis (mapping, if any) of activities under both active and passive surveillance (to contribute identify possible gaps in disease surveillance).

All sampling events must be validated and registered by DGAV local and regional services. At central level, records for each sampling event, based on analytical data sent by INIAV are also kept and monthly checks are carried out to assess the operationalisation of the programme.

2.2 Programme participants (stakeholders)

Cooperation and division of roles and responsibilities

Indicate participants (stakeholders such as competent authorities, testing laboratories, authorised private veterinarians other stakeholders as relevant) involved in the planning and implementation of the programme; what are their roles and responsibilities; who reports to whom; what are the reporting arrangements.

Indicate who is overall responsible for the programme and how the overall responsible coordinates with other stakeholders; how effective communication will be ensured.

2.2.1 - Poultry

The General Directorate of Food and Veterinary (DGAV) is responsible for the implementation of programme, defining objectives, strategies and action guidelines as well as coordinating all those involved on its execution in each region.

At central level, Epidemiology and Animal Health Unit (DESA) oversees designing, supervising, monitoring and evaluating the programme. DESA also carries out data collection and analysis and submits the programme's annual and semi-annual reports to the European Commission and EFSA, respectively.

At regional level, the local Food and Veterinary Regional Departments of DGAV (DSAVR) on the Mainland or the Regional Directions of Agriculture (DRA) in the Autonomous Regions of Madeira and Açores implement the programme, carrying out sampling and delivering samples to the laboratory, National Institute of Agrarian and Veterinary Research (INIAV).

All laboratory testing is carried out by INIAV, which is the national reference laboratory (NRL) for avian influenza and results are regularly sent to DGAV.

There are regular contacts between DESA and DSAVR/DRA for the purpose of correcting any deviations to the targets established in the programme and for assessing any difficulties of local services regarding the programme implementation in the field.

Whenever changes to the number of existing holdings justify it, the number of holdings to be testing is also updated by DESA.

2.2.2 – Wild birds

As for poultry, DGAV is responsible for the programme, defining the objectives, strategies and action guidelines and coordinating involved people and organizations on the actions to be implemented in each region. At central level, DESA will establish, supervise, monitor, and evaluate the programme. The collection of samples from wild birds, coordinated regionally by the DSAVR/DRA is performed mostly by official veterinarians of the local veterinary services, field workers of the Institute for Nature Conservation and Forestry, IP (ICNF), by special brigades of the National Republican Guard and Police Force and by the municipality veterinarians. Nature conservation organizations, bird ringing teams, hunters and ornithologists may also collaborate in the sample collection.

It is the responsibility of the DSAVR/DRA to articulate with other entities in the field to ensure the correct implementation of the programme, delivery of the samples to INIAV and the correct filling of the request forms that accompany the samples. Control and supervision procedures are the same as described for poultry.

All mass mortality events of wild birds must be notified to DGAV, either at central or local level. Personnel of the organizations involved (please see above) regularly carries out routine rounds in protected natural areas and in rural areas where dead wild birds can be found. Also, wounded, diseased and or dead wild birds entering wild bird rescue centres can also be sampled. An app called ANIMAS, developed by DGAV and ICNF, for immediate notification of wild animals found dead in natural settings, is publicly available. When dead wild birds are reported through this app, a risk assessment regarding the number, species and location of the event is done and sampling is carried out if necessary.

Upon detection of dead or injured wild birds, local DSAVR/DRA services are contacted, and sampling is carried out by official veterinarians. The DSAVR/DRA services are responsible for the delivery of samples to INIAV, I.P, the national reference laboratory for animal health, where testing is performed.

2.3 Management; controls and verifications, quality assurance and monitoring and evaluation strategy

Describe the activities planned to ensure that the implementation of the programme activities is of high quality and completed in time (according to the plan/timeline). Explain planned controls and verifications, and monitoring of the achievement of targets (activity[1] indicators) - please describe for different programme activities; mention frequency of such controls.

What enforcement mechanisms will be initiated in case of failure of reaching the planned targets / to ensure continuous improvement.

Describe the evaluation of the progress[2] indicators (quantitative or qualitative); the outreach of the expected results/outcome (include unit of measurement, baseline and target values). The indicators proposed to measure progress (progress indicators) should be relevant, realistic, and measurable.

Monthly, DESA collects, completes, verifies, and validates data (request forms and analytical results) received from INIAV in a database and monitors the implementation of the surveillance programme. The average timing of samples' delivery to the laboratory, the laboratory average response timing and the quality of the information that complements the sample (request forms) are also assessed by DESA. Additionally, DESA verifies and validates the monthly technical information and invoices sent by INIAV, regarding the number of tests carried out under the programme in the mainland.

The implementation of the programme is assessed through the number of sampling events carried out by DGAV local services.

2.4 Risk management

[1] Example of activity indictors: number of holdings checked; number of animals samples; number of samples tested, etc.

[2] Example of progress indicators: number of samples tested under passive surveillance higher than the last year, indicating higher likelihood of early detection of possible introduction of disease (new disease outbreaks).

Critical risks and risk management strategy

Describe critical risks, uncertainties or difficulties related to the implementation of the programme, and mitigation measures/strategy for addressing them.

Indicate for each risk (in the description) the impact and the likelihood that the risk will materialise (high, medium, low), even after taking into account the mitigating measures.

***Note:** Uncertainties and unexpected events occur in all organizations, even if very well-run. The risk analysis will help you to predict issues that could delay or hinder project activities. A good risk management strategy is essential for good project management.*

Risk No	Description	Proposed risk-mitigation measures
1	Deadlines for contracting laboratory services – public procurement	Timely launch of contractual procedures to start analytics

2.5 Milestones

Indicate control points along the programme implementation that help to chart progress.

Note: Deliverables (*e.g. intermediate or final report on the implementation of programme measures*) are not milestones.

Name	Due date (in month)	Means of verification
Verification of sampling events	monthly	Collection and analysis of laboratory results

3. IMPACT

3.1 Impact and ambition

*Describe **expected impact** (benefit) of the programme (e.g. from the economical and animal health points of view).*

Who are the target groups? How will the target groups benefit concretely from the project and what would change for them?

Define the short, medium and long-term effects of the project.

***Possible examples**: increased likelihood of early detection and response in case of disease occurrence, contributes decrease in preventable losses in animal production and loses due to trade restrictions.*

Given the current epidemiological situation of avian influenza, namely highly pathogenic AI, in Europe, this programme is key for achieving a favourable evolution regarding this disease through:

a) Monitoring of virus circulation in wild birds.
b) An early detection and a rapid and effective response in case of outbreak's confirmation.
c) A higher level of protection of poultry health and welfare.
d) Reduction of economic losses due to killing and destruction of infected flocks and its products as well as trade restrictions.
e) Protecting public health and avoiding disruption in the food chain.
f) Protecting biodiversity by identification and safe disposal of infected wild birds to break disease transmission and minimise environmental contamination.

3.2 Communication, dissemination and visibility

Communication, dissemination and visibility of funding

Describe the communication and information dissemination activities which are planned in order to promote the activities/results and maximise the impact (to whom, which format, how many, etc.).

Describe how the visibility of EU funding will be ensured.

Coordination meetings are held to present the programme to local veterinary services which are responsible for sampling as well as to discuss the progress of the implementation of the programme.

Information about this programme is publicly available at DGAV web portal. There is an Avian Influenza dedicated page in this portal containing information about:

a) Disease detection, diagnosis, and notification of suspicions.
b) Epidemiological situation of avian influenza in Portugal, including outbreaks data and control and eradication measures enforced, in Europe and in the rest of the world.
c) Information for poultry keepers regarding biosecurity and other relevant issues (compensations, movement restrictions and derogations).

Regular awareness sessions about avian influenza for poultry industry stakeholders, including epidemiological situation, prevention, and biosecurity as well as activities within the scope of the surveillance programme, are held either online or physically.

3.3 Sustainability and continuation

Sustainability, long-term impact and continuation *Describe the how will the project impact be ensured and sustained long term? Which parts of the project should be continued or maintained, and which resources will be necessary to continue?* *Are there any possible synergies/complementarities with other (EU funded) activities that can build on the results of the implementation of this project?*
Avian Influenza, namely its highly pathogenic form, is currently a major animal health problem worldwide. An efficient surveillance programme is essential for early detection and timely implementation of outbreak control and eradication measures. Outbreak ‘s prevention and control rely on appropriate biosecurity measures, surveillance, and rapid eradication. Hence, surveillance, in wild and kept birds, is key for controlling this zoonosis.

ANNEX

I. **Baseline population data and Targets for 2024**
II. **History of disease occurrence**
III. **Implementation of applicable rules and regulation**
IV. **Maps (as relevant)**
V. **Wild birds population data**

I. Baseline population data and targets for 2024

Table 1a: Poultry holdings[3] (except ducks, geese and farmed game birds (waterfowl eg. Mallards) to be sampled (insert as appropriate for the programme)

Laying hens

Number	Region (NUTS-2)[1]	Total number of holdings[2]	Total number of holdings to be sampled	Number of samples per holding	Total number of samples	Total number of tests	Method of laboratory analysis[3]
403 030	PT11-Norte	10	4	20	80	80	ELISA
4 010 296	PT16-Centro	70	34	20	680	680	ELISA
2 515 936	PT17-Lisboa e Vale do Tejo	22	19	20	380	380	ELISA
28 000	PT18-Alentejo	1	1	20	20	20	ELISA
0	PT15-Algarve	0	0	20	0	0	ELISA
201822	PT20-Açores	8	0	20	0	0	ELISA
108 088	PT30-Madeira	5	0	20	0	0	ELISA
	All country					23	HI test (H5)
	All country					23	HI test (H7)
	TOTAL	116	58		1160	1206	

[2] Holdings or herds or flocks or establishments as appropriate.

Free range laying hens

Number	Region (NUTS-2)[1]	Total number of holdings[2]	Total number of holdings to be sampled	Number of samples per holding	Total number of samples	Total number of tests	Method of laboratory analysis[3]
103 148	PT11-Norte	4	3	20	60	60	ELISA
526 142	PT16-Centro	29	27	20	540	540	ELISA
17 700	PT17-Lisboa e Vale do Tejo	6	6	20	120	120	ELISA
4 423	PT18-Alentejo	5	4	20	80	80	ELISA
0	PT15-Algarve	0	0	20	0	0	ELISA
0	PT20-Açores	0	0	20	0	0	ELISA
110	PT30-Madeira	2	0	20	0	0	ELISA
	All country					16	HI - H5
	All country					16	HI - H7
	TOTAL	46	40		800	832	

Fattening turkeys

Number	Region (NUTS-2)[1]	Total number of holdings[2]	Total number of holdings to be sampled	Number of samples per holding	Total number of samples	Total number of tests	Method of laboratory analysis[3]
30 000	PT11-Norte	1	0	10	0	0	ELISA
302 459	PT16-Centro	39	20	10	200	200	ELISA
1 567 481	PT17-Lisboa e Vale do Tejo	98	45	10	450	450	ELISA
60 000	PT18-Alentejo	3	2	10	20	20	ELISA
0	PT15-Algarve	0	0	10	0	0	ELISA
0	PT20-Açores	0	0	10	0	0	ELISA
0	PT30-Madeira	0	0	10	0	0	ELISA
	All country					13	HI - H5
	All country					13	HI - H7
	TOTAL	141	67		670	696	

Farmed game birds (gallinaceous)

Number	Region (NUTS-2)[1]	Total number of holdings[2]	Total number of holdings to be sampled	Number of samples per holding	Total number of samples	Total number of tests	Method of laboratory analysis[3]
13 082	PT11-Norte	11	11	10	110	22	PCR
3 181	PT16-Centro	6	6	10	60	12	PCR
13 270	PT17-Lisboa e Vale do Tejo	9	9	10	90	18	PCR
9 000	PT18-Alentejo	3	3	10	30	6	PCR
1 560	PT15-Algarve	2	2	10	20	4	PCR
0	PT20-Açores	0	0	10	0	0	PCR
1 200	PT30-Madeira	1	1	10	10	2	PCR
	TOTAL	32	32		320	64	

Other please specify here: Quails

Number	Region (NUTS-2)[1]	Total number of holdings[2]	Total number of holdings to be sampled	Number of samples per holding	Total number of samples	Total number of tests	Method of laboratory analysis[3]
0	PT11-Norte	0	0	10	0	0	PCR
39 000	PT16-Centro	1	1	10	10	2	PCR
1 717 680	PT17-Lisboa e Vale do Tejo	16	16	10	160	32	PCR
0	PT18-Alentejo	0	0	10	0	0	PCR
0	PT15-Algarve	0	0	10	0	0	PCR
0	PT20-Açores	0	0	10	0	0	PCR
1 750	PT30-Madeira	1	1	10	10	2	PCR
	TOTAL	18	18		180	36	

Totals	Total number of tests	Total number of samples
Total poultry Galliformes 2024	2834	3130

In the column "Total number of samples", please put 0 if the same samples have already been counted for another laboratory analysis (example : for HI-H5 and HI –H7 test, only 1 sample should be counted)

Table 1b: DUCKS, GEESE AND FARMED GAME BIRDS (WATERFOWL eg. MALLARD) HOLDING[4] to be sampled

(insert as appropriate for the programme)

Duck breeders

Number	Region (NUTS-2)[1]	Total number of holdings[2]	Total number of holdings to be sampled	Number of samples per holding	Total number of samples	Total number of tests	Method of laboratory analysis[3]
0	PT11-Norte	0	0	40	0	0	PCR
0	PT16-Centro	0	0	40	0	0	PCR
65 280	PT17-Lisboa e Vale do Tejo	2	2	40	80	16	PCR
0	PT18-Alentejo	0	0	40	0	0	PCR
0	PT15-Algarve	0	0	40	0	0	PCR
0	PT20-Açores	0	0	40	0	0	PCR
0	PT30-Madeira	0	0	40	0	0	PCR
	TOTAL	2	2		80	16	

[4] Holdings or herds or flocks or establishments as appropriate.

Fattening ducks

Number	Region (NUTS-2)[1]	Total number of holdings[2]	Total number of holdings to be sampled	Number of samples per holding	Total number of samples	Total number of tests	Method of laboratory analysis[3]
0	PT11-Norte	0	0	60	0	0	PCR
42 600	PT16-Centro	3	3	60	180	36	PCR
777 972	PT17-Lisboa e Vale do Tejo	15	15	60	900	180	PCR
76 062	PT18-Alentejo	2	2	60	120	24	PCR
0	PT15-Algarve	0	0	60	0	0	PCR
0	PT20-Açores	0	0	60	0	0	PCR
0	PT30-Madeira	0	0	60	0	0	PCR
	TOTAL	20	20		1200	240	

Farmed game (waterfowl eg. Mallards)

Number	Region (NUTS-2)[1]	Total number of holdings[2]	Total number of holdings to be sampled	Number of samples per holding	Total number of samples	Total number of tests	Method of laboratory analysis[3]
200	PT11-Norte	1	1	40	40	8	PCR
0	PT16-Centro	0	0	40	0	0	PCR
4 200	PT17-Lisboa e Vale do Tejo	1	1	40	40	8	PCR
0	PT18-Alentejo	0	0	40	0	0	PCR
0	PT15-Algarve	0	0	40	0	0	PCR
0	PT20-Açores	0	0	40	0	0	PCR
0	PT30-Madeira	0	0	40	0	0	PCR
	TOTAL	2	2		80	16	

Totals	Total number of tests	Total number of samples
Total ducks and geese and farmed waterfowl game birds 2024	272	1360

TOTALS for Poultry (Table 1a) & Ducks and Geese (Table 1b) and farmed game birds for year: 2024

Poultry & Ducks/Geese/farmed game birds	Total number of tests
Grand Total	3106
Grand Total ELISA	2630
Grand Total Agar	0
Grand Total HI tests (H5)	52
Grant Total HI tests (H7)	52
Grand Total Virus Isolation test	0
Grand Total PCR test	372
Grand Total Sampling	4490

Table 1c: WILD BIRDS focussed on target species

Targets for year: 2024

Number	Region (NUTS-2)[1]	Total number of wild birds to be sampled	Estimated total number of wild birds to be sampled for passive surveillance	Type of test	Number of tests
All countryPlease see point 1.4.3 and annex V	PT11-Norte	20	20	real-time RT-PCR	20
	PT16-Centro	40	40	real-time RT-PCR	40
	PT17-Lisboa e Vale do Tejo	40	40	real-time RT-PCR	40
	PT18-Alentejo	30	30	real-time RT-PCR	30
	PT15-Algarve	60	60	real-time RT-PCR	60
	PT20-Açores	15	15	real-time RT-PCR	15
	PT30-Madeira	15	15	real-time RT-PCR	15
	All country			Virus isolation test	15
	TOTAL	220	220		220

	Total number of tests
Total number of tests	**220**
Total Virus isolation tests	**15**
Total PCR tests	**220**
Total Other tests	**15**
Total number of wild birds to be sampled for passive surveillance	**220**

II. History of disease outbreaks

Table 2: Poultry and wild birds' outbreaks

Outbreaks				
Year	Poultry establishments [1]		Wild birds (nº)	
	Checked	Positive	Examined	Positive
2018	372	0	83	0
2019	352	0	84	0
2020	324	0	74	0
2021	391	3	64	0
2022	359	16	221[3]	18[2]

[1] Including active and passive surveillance and outbreak related reinforced surveillance in protection and surveillance zones in commercial poultry establishments and non-commercial backyard flocks.

[2] Corresponding to 13 outbreaks.

[3] Including 40 wild birds tested in wildlife rescue centres before their release into nature, as part of reinforced surveillance implemented due to HPAI outbreaks.

III. Implementation of applicable rules and regulation

(TRACEABILITY, DISEASE NOTIFICATION AND MEASURES FOR EFFECTIVE DETECTION AND ELIMINATION OF THE DISEASE)

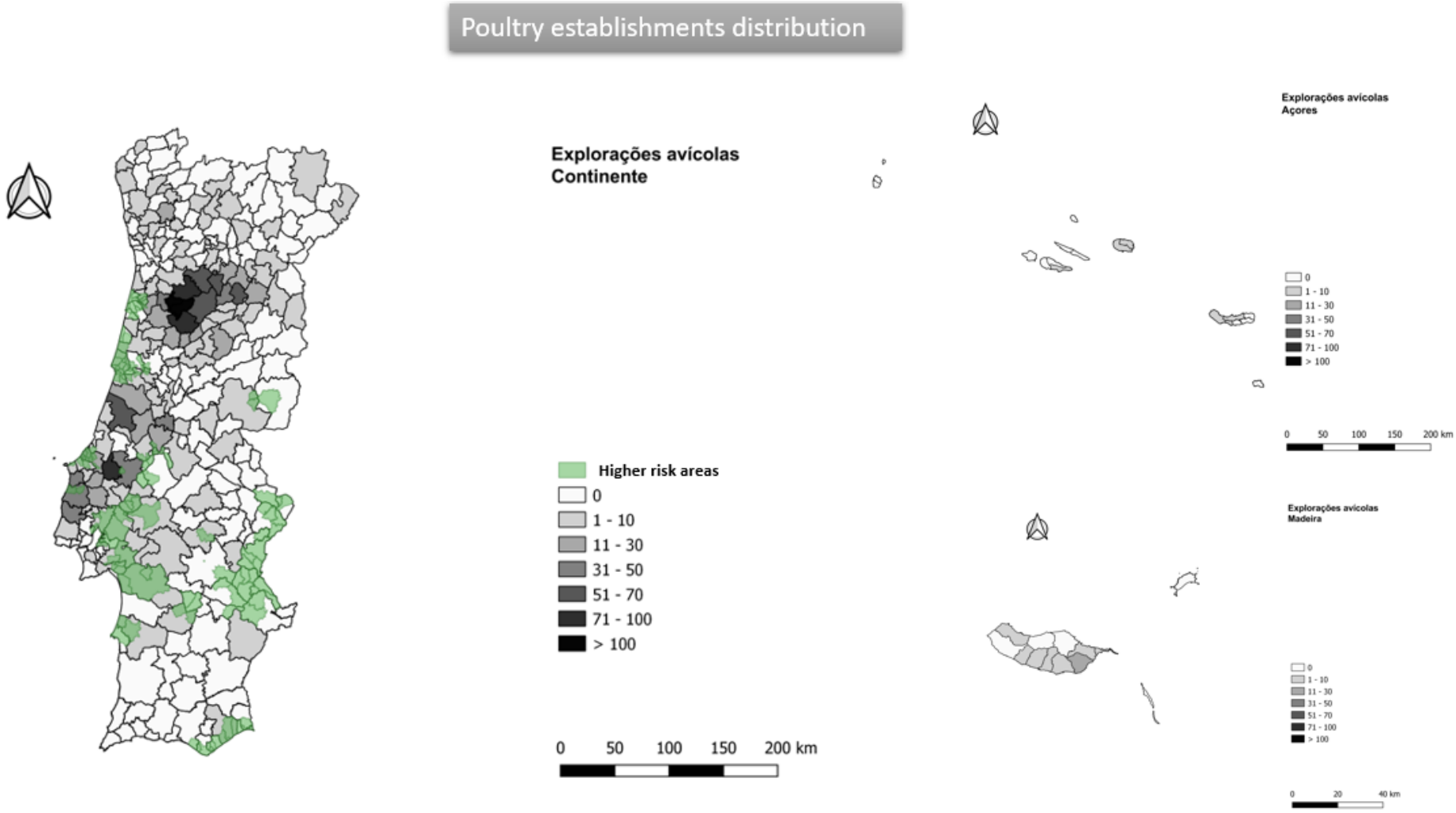
EU countries

Implementation of applicable regulations - please tick the box as appropriate. In case of deviations, please describe / justify.

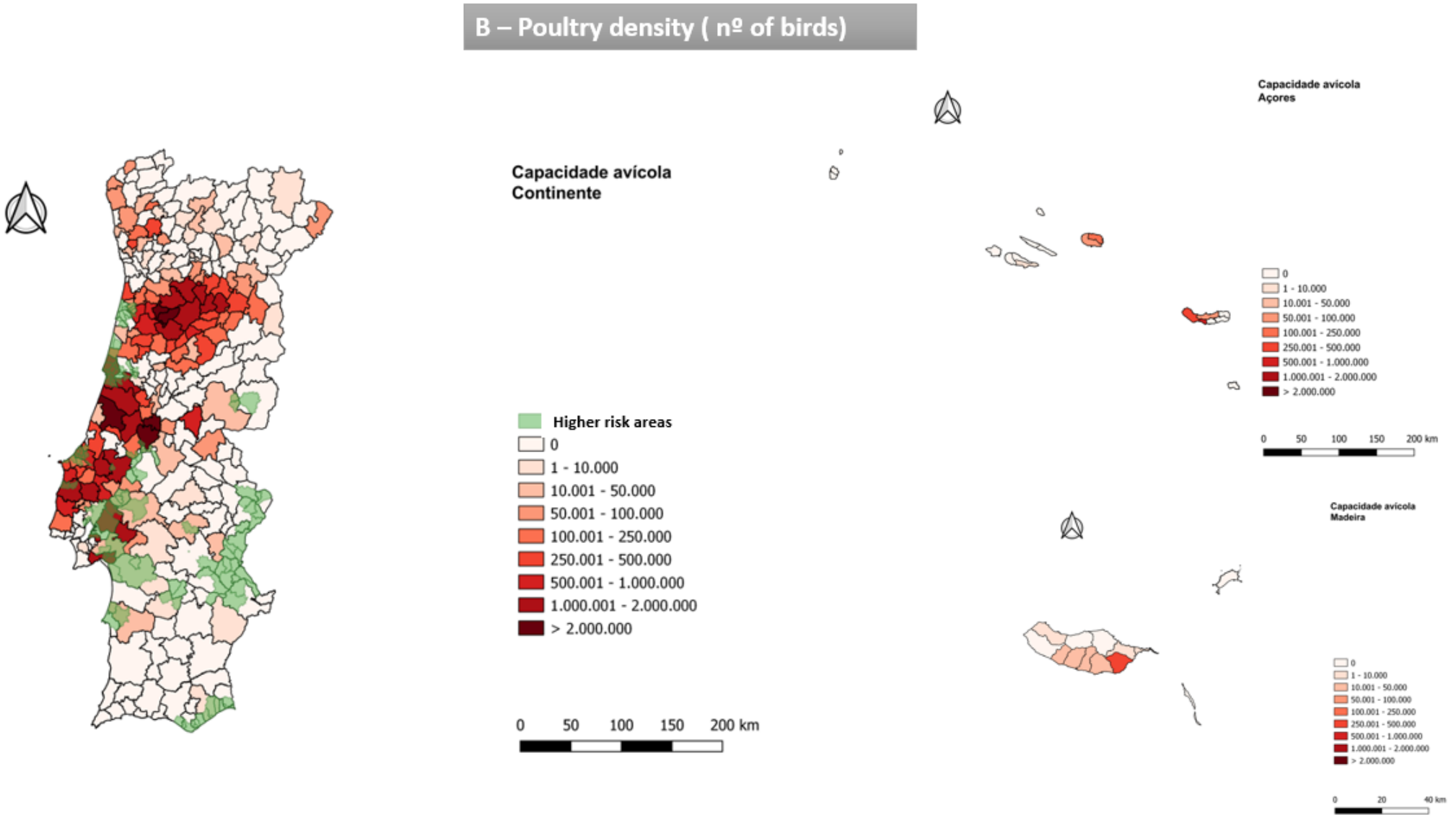
1.	**Regulation (EU) 2016/429** of the European Parliament and of the Council of 9 March 2016 on transmissible animal diseases and amending and repealing certain acts in the area of animal health ('Animal Health Law') (OJ L 84, 31.3.2016, pp. 1-208)	**Yes** ☒ **No** ☐	*Description of deviation/Justification (when relevant):*
2.	**Commission Delegated Regulation (EU) 2020/687** of 17 December 2019 supplementing Regulation (EU) 2016/429 of the European Parliament and the Council, as regards rules for the prevention and control of certain listed diseases (OJ L 174, 3.6.2020, pp. 64-139).	**Yes** ☒ **No** ☐	*Description of deviation/Justification (when relevant):*
3.	**Commission Implementing Regulation (EU) 2020/2002** of 7 December 2020 laying down rules for the application of Regulation (EU) 2016/429 of the European Parliament and of the Council with regard to Union notification and Union reporting of listed diseases, to formats and procedures for submission and reporting of Union surveillance programmes and of eradication programmes and for application for recognition of disease-free status, and to the computerised information system (OJ L 412, 8.12.2020, pp. 1-28).	**Yes** ☒ **No** ☐	*Description of deviation/Justification (when relevant):*
4.	**Commission Implementing Regulation (EU) 2020/690** of 17 December 2019 laying down rules for the application of Regulation (EU) 2016/429 of the European Parliament and of the Council as regards the listed diseases subject to Union surveillance programmes, the geographical scope of such programmes and the listed diseases for which the disease-free status of compartments may be established (OJ L 174, 3.6.2020, pp. 341-344)	**Yes** ☒ **No** ☐	*Description of deviation/Justification (when relevant):*
5.	**Commission Delegated Regulation (EU) 2020/689** on 17 December 2019 supplementing Regulation (EU) 2016/429 of the European Parliament and of the Council as regards rules for surveillance, eradication programmes, and disease-free status for certain listed and emerging diseases **(**OJ L 174, 3.6.2020, p. 211–340)	**Yes** ☒ **No** ☐	*Description of deviation/Justification (when relevant):*

IV. Maps (as relevant)

A - POULTRY ESTABLISHMENTS DISTRIBUTION



B - POULTRY DENSITY



V. Wild birds population data

Annex I: Article 12 National Summary Factsheet – Portugal

List of bird species reported by the Member State with the overall short-term and long-term population trends for the reporting period 2013-2018.

PORTUGAL - MAINLAND

Regularly occurring bird species (PRE). These are included in the statistics of the National Summary.

species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend	
A899	Accipiter gentilis all others	breeding	N	100	500	NULL	p	no change	x	x	
A898	Accipiter nisus all others	breeding	N	200	1300	NULL	p	no change	-	x	
A298	Acrocephalus arundinaceus	breeding	N	600	2500	NULL	p	different method	u	x	
A297	Acrocephalus scirpaceus	breeding	N	1000	5000	NULL	p	different method	x	x	
A168	Actitis hypoleucos	breeding	N	500	1000	NULL	p	no change	u	x	
A324	Aegithalos caudatus	breeding	N	500000	1000000	NULL	p	no change	-	x	
A247	Alauda arvensis	breeding	N	10000	50000	NULL	p	different method	x	x	
A475	Alaudala rufescens	breeding	N	10	100	NULL	p	no change	x	x	
A229	Alcedo atthis	breeding	Y	5000	30000	NULL	p	different method	-	x	
A110	Alectoris rufa	breeding	N	500000	1000000	NULL	p	no change	-	x	
A053	Anas platyrhynchos	breeding	N	100000	200000	NULL	p		+	+	
A255	Anthus campestris	breeding	Y	1000	10000	NULL	p	no change	-	x	
A259	Anthus spinoletta	breeding	N	10	50	NULL	p	no change	x	x	
A256	Anthus trivialis	breeding	N	100	1000	NULL	p	no change	-	x	
A226	Apus apus	breeding	N	100000	1000000	NULL	p		-	x	
A424	Apus caffer	breeding	Y	50	100	NULL	p	genuine change	x	x	
A227	Apus pallidus	breeding	N	100000	1000000	NULL	p		u	x	
A405	Aquila adalberti	breeding	Y	17	17	17	p	no change	+	N/A	
A091	Aquila chrysaetos	breeding	Y	70	85	NULL	p	no change	x	x	

In the table above 'N/A' means no data were provided, 'NULL' means information was not expected.

species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend	
A707	Aquila fasciata	breeding	Y	150	180	NULL	**p**	no change	x	x	
A028	Ardea cinerea	breeding	N	1234	1550	NULL	**p**	no change	=	=	
A029	Ardea purpurea	breeding	Y	797	920	NULL	**p**	no change	+	+	
A024	Ardeola ralloides	breeding	Y	10	50	NULL	**p**	no change	x	x	
A221	Asio otus	breeding	N	100	500	NULL	**p**	different method	-	x	
A218	Athene noctua	breeding	N	40000	60000	NULL	**p**	different method	-	x	
A059	Aythya ferina	breeding	N	30	100	NULL	**p**	different method	x	x	
A060	Aythya nyroca	breeding	Y	10	25	NULL	**p**	different method	x	x	
A021	Botaurus stellaris	breeding	Y	1	1	NULL	**cmales**	no change	x	x	
A215	Bubo bubo	breeding	Y	150	500	NULL	**p**	no change	u	x	
A025	Bubulcus ibis	breeding	N	14900	18000	NULL	**p**	no change	+	x	
A133	Burhinus oedicnemus	breeding	Y	500	5000	NULL	**p**	no change	u	x	
A087	Buteo buteo	breeding	N	5000	10000	NULL	**p**	no change	-	x	
A243	Calandrella brachydactyla	breeding	Y	5000	10000	NULL	**p**	different method	-	x	
A851	Calonectris borealis	breeding	Y	800	975	NULL	**p**	no change	u	+	
A224	Caprimulgus europaeus	breeding	Y	1000	5000	NULL	**cmales**	different method	+	x	
A225	Caprimulgus ruficollis	breeding	N	1000	5000	NULL	**cmales**	different method	u	x	
A364	Carduelis carduelis	breeding	N	500000	2000000	NULL	**p**	different method	-	x	
A479	Cecropis daurica	breeding	N	10000	100000	NULL	**p**	genuine change	+	+	
A268	Cercotrichas galactotes	breeding	N	200	500	NULL	**p**	genuine change	-	x	
A637	Certhia brachydactyla all others	breeding	N	500000	1000000	NULL	**p**	no change	=	=	
A288	Cettia cetti	breeding	N	10000	50000	NULL	**p**	different method	+	=	

In the table above 'N/A' means no data were provided, 'NULL' means information was not expected.

species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend	
A138	Charadrius alexandrinus	breeding	Y	1000	5000	NULL	**p**	no change	=	x	
A136	Charadrius dubius	breeding	N	1000	5000	NULL	**p**	no change	u	x	
A734	Chlidonias hybrida	breeding	Y	5	50	NULL	**p**	no change	x	x	
A363	Chloris chloris	breeding	N	500000	1000000	NULL	**p**	no change	-	x	
A031-A	Ciconia ciconia - W Europe & North-west Africa/Sub-Saharan Africa	breeding	Y	12400	12600	12307	**p**	genuine change	=	+	
A030	Ciconia nigra	breeding	Y	100	150	NULL	**p**	no change	-	x	
A264	Cinclus cinclus	breeding	N	500	2500	NULL	**p**	different method	-	x	
A080	Circaetus gallicus	breeding	Y	500	1000	NULL	**p**	no change	x	x	
A081	Circus aeruginosus	breeding	Y	150	300	NULL	**bfemales**	no change	x	x	
A082	Circus cyaneus	breeding	Y	20	50	NULL	**bfemales**	no change	x	x	
A084	Circus pygargus	breeding	Y	300	1000	NULL	**bfemales**	no change	x	x	
A289	Cisticola juncidis	breeding	N	100000	500000	NULL	**p**	no change	=	=	
A211	Clamator glandarius	breeding	N	1000	5000	NULL	**p**	no change	-	x	
A373	Coccothraustes coccothraustes	breeding	N	5000	10000	NULL	**p**	no change	x	x	
A206-X	Columba livia - feral populations	breeding	N	5000000	8000000	NULL	**p**		+	x	
A206	Columba livia - wild populations	breeding	N	NULL	NULL	NULL	**NULL**		N/A	N/A	
A207	Columba oenas	breeding	N	200	1000	NULL	**p**	different method	u	x	
A687	Columba palumbus palumbus	breeding	N	100000	500000	NULL	**p**	no change	+	x	
A231	Coracias garrulus	breeding	Y	50	120	NULL	**p**	no change	x	x	
A350	Corvus corax	breeding	N	500	1500	NULL	**p**	no change	-	x	
A349	Corvus corone - [including cornix]	breeding	N	100000	500000	NULL	**p**	no change	=	x	

In the table above 'N/A' means no data were provided, 'NULL' means information was not expected.

species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend	
A347	Corvus monedula	breeding	N	1000	5000	NULL	**p**	no change	u	x	
A113	Coturnix coturnix	breeding	N	50000	100000	NULL	**cmales**	different method	=	=	
A212	Cuculus canorus	breeding	N	50000	100000	NULL	**cmales**	no change	-	x	
A483	Cyanistes caeruleus s. str. - sensu stricto [excluding teneriffae]	breeding	N	1000000	5000000	NULL	**p**	no change	=	=	
A481	Cyanopica cooki	breeding	N	100000	500000	NULL	**p**	no change	+	+	
A738	Delichon urbicum	breeding	N	500000	2000000	NULL	**p**	different method	=	=	
A658	Dendrocopos major all others	breeding	N	50000	150000	NULL	**p**	different method	=	=	
A869	Dryobates minor	breeding	N	2500	5000	NULL	**p**	different method	u	x	
A026	Egretta garzetta	breeding	Y	1031	1600	NULL	**p**	no change	x	x	
A399	Elanus caeruleus	breeding	Y	600	1500	NULL	**p**	no change	u	x	
A383	Emberiza calandra	breeding	N	500000	1000000	NULL	**p**	no change	=	=	
A378	Emberiza cia	breeding	N	100000	500000	NULL	**p**	no change	-	x	
A377	Emberiza cirlus	breeding	N	100000	500000	NULL	**p**	no change	=	=	
A376	Emberiza citrinella	breeding	N	100	250	NULL	**p**	different method	u	x	
A379	Emberiza hortulana	breeding	Y	500	2500	NULL	**p**	different method	x	x	
A381	Emberiza schoeniclus	breeding	N	50	300	NULL	**p**	different method	u	x	
A269	Erithacus rubecula	breeding	N	500000	1000000	NULL	**p**	no change	+	=	
A095	Falco naumanni	breeding	Y	674	735	NULL	**p**	genuine change	+	+	
A103	Falco peregrinus - [including pelegrinoides]	breeding	Y	100	200	NULL	**p**	no change	x	x	
A099	Falco subbuteo	breeding	N	100	250	NULL	**p**	different method	x	x	
A096	Falco tinnunculus	breeding	N	1500	3000	NULL	**p**	no change	=	x	

In the table above ‘N/A’ means no data were provided, ‘NULL’ means information was not expected.

species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend	
A657	Fringilla coelebs all others	breeding	N	1000000	5000000	NULL	p	no change	=	=	
A125	Fulica atra	breeding	N	4000	20000	NULL	p		+	+	
A244	Galerida cristata	breeding	N	100000	500000	NULL	p	no change	+	+	
A245	Galerida theklae	breeding	Y	100000	500000	NULL	p	no change	-	x	
A153	Gallinago gallinago	breeding	N	1	5	NULL	p	no change	x	x	
A123	Gallinula chloropus	breeding	N	5000	25000	NULL	p		u	x	
A342	Garrulus glandarius	breeding	N	100000	500000	NULL	p	no change	=	x	
A189	Gelochelidon nilotica	breeding	Y	1000	2000	NULL	p	no change	x	x	
A135	Glareola pratincola	breeding	Y	500	1000	NULL	p	no change	x	x	
A078	Gyps fulvus	breeding	Y	500	1000	NULL	p	no change	x	x	
A092	Hieraaetus pennatus	breeding	Y	1500	4000	NULL	p	no change	x	x	
A131	Himantopus himantopus	breeding	Y	2000	5000	NULL	p	no change	+	+	
A300	Hippolais polyglotta	breeding	N	10000	500000	NULL	p	no change	=	x	
A251	Hirundo rustica	breeding	N	500000	1000000	NULL	p	different method	-	x	
A874	Hydrobates castro	breeding	Y	410	784	NULL	p	genuine change	x	x	
A022	Ixobrychus minutus	breeding	Y	100	500	NULL	p	different method	x	x	
A233	Jynx torquilla	breeding	N	1000	5000	NULL	p	no change	u	x	
A338	Lanius collurio	breeding	Y	100	500	NULL	p	no change	u	x	
A496	Lanius meridionalis - [excluding koenigi]	breeding	N	1000	10000	NULL	p	genuine change	-	-	
A341	Lanius senator	breeding	N	10000	50000	NULL	p	genuine change	-	-	
A181	Larus audouinii	breeding	Y	1900	2000	NULL	p	genuine change	x	x	

In the table above ‘N/A’ means no data were provided, ‘NULL’ means information was not expected.

species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend	
A489	Larus fuscus all others	breeding	N	20	50	NULL	p	no change	x	x	
A604	Larus michahellis	breeding	N	15000	40000	NULL	p	no change	x	x	
A179	Larus ridibundus	breeding	N	10	40	NULL	p	genuine change	x	x	
A476	Linaria cannabina	breeding	N	500000	1000000	NULL	p	no change	=	x	
A292	Locustella luscinioides	breeding	N	1000	5000	NULL	p	no change	u	x	
A497	Lophophanes cristatus	breeding	N	100000	500000	NULL	p	no change	u	x	
A369	Loxia curvirostra	breeding	N	100	500	NULL	p	no change	x	x	
A246	Lullula arborea	breeding	Y	100000	500000	NULL	p	no change	-	x	
A271	Luscinia megarhynchos	breeding	N	100000	500000	NULL	p	different method	=	=	
A889	Mareca strepera	breeding	N	500	4000	NULL	p	different method	+	+	
A242	Melanocorypha calandra	breeding	Y	5000	10000	NULL	p	no change	u	x	
A230	Merops apiaster	breeding	N	100000	500000	NULL	p		-	-	
A073	Milvus migrans	breeding	Y	1000	2000	NULL	p	no change	x	x	
A074	Milvus milvus	breeding	Y	50	100	NULL	p	no change	x	x	
A280	Monticola saxatilis	breeding	N	100	500	NULL	p	no change	x	x	
A281	Monticola solitarius	breeding	N	1000	5000	NULL	p	no change	u	u	
A262	Motacilla alba	breeding	N	100000	500000	NULL	p	no change	+	+	
A261	Motacilla cinerea	breeding	N	10000	50000	NULL	p	no change	=	x	
A260	Motacilla flava - [excluding tschutschensis]	breeding	N	1000	5000	NULL	p	no change	u	x	
A319	Muscicapa striata	breeding	N	500	1000	NULL	p	no change	u	x	
A077	Neophron percnopterus	breeding	Y	45	90	NULL	p	no change	x	x	
A058	Netta rufina	breeding	N	500	1000	NULL	p	different method	+	+	

In the table above ‘N/A’ means no data were provided, ‘NULL’ means information was not expected.

species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend	
A023	Nycticorax nycticorax	breeding	Y	244	300	NULL	p	no change	x	x	
A278	Oenanthe hispanica	breeding	N	1000	5000	NULL	p	genuine change	x	x	
A279	Oenanthe leucura	breeding	Y	50	100	NULL	p	genuine change	x	x	
A277	Oenanthe oenanthe - [excluding seebohmi]	breeding	N	1000	5000	NULL	p	genuine change	-	x	
A337	Oriolus oriolus	breeding	N	10000	100000	NULL	p	different method	=	=	
A129	Otis tarda	breeding	Y	320	320	NULL	i	genuine change	-	-	
A214	Otus scops	breeding	N	1000	5000	NULL	p	different method	-	x	
A330	Parus major	breeding	N	1000000	5000000	NULL	p	no change	-	x	
A620	Passer domesticus s. str. - sensu stricto [excluding italiae]	breeding	N	1000000	3000000	NULL	p	different method	-	x	
A355	Passer hispaniolensis	breeding	N	50000	500000	NULL	p	different method	+	x	
A356	Passer montanus	breeding	N	100000	500000	NULL	p	no change	-	x	
A472	Periparus ater all others	breeding	N	100000	500000	NULL	p	no change	-	x	
A072	Pernis apivorus	breeding	Y	150	300	NULL	p	no change	x	x	
A357	Petronia petronia	breeding	N	50000	100000	NULL	p	no change	x	x	
A684	Phalacrocorax aristotelis aristotelis	breeding	N	100	150	NULL	p	no change	x	x	
A115-X	Phasianus colchicus - non-native populations	breeding	N	10	90	NULL	p	different method	x	x	
A273	Phoenicurus ochruros	breeding	N	50000	100000	NULL	p	no change	+	+	
A274	Phoenicurus phoenicurus	breeding	N	1000	5000	NULL	p	no change	u	x	
A499	Phylloscopus bonelli s. str. - sensu stricto [excluding orientalis]	breeding	N	10000	50000	NULL	p	no change	=	x	
A618	Phylloscopus ibericus	breeding	N	50000	100000	NULL	p	no change	+	x	

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species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend	
A343	Pica pica	breeding	N	100000	500000	NULL	p	no change	+	+	
A867	Picus sharpei	breeding	N	10000	50000	NULL	p	no change	u	x	
A034	Platalea leucorodia	breeding	Y	518	550	NULL	p	genuine change	+	x	
A005	Podiceps cristatus	breeding	N	600	5000	NULL	p	different method	u	x	
A722	Porphyrio porphyrio porphyrio	breeding	Y	100	400	NULL	p	genuine change	x	x	
A266	Prunella modularis	breeding	N	25000	50000	NULL	p	different method	-	x	
A205	Pterocles alchata	breeding	Y	3	16	NULL	p	no change	x	x	
A420	Pterocles orientalis	breeding	Y	130	150	NULL	p	no change	-	-	
A250	Ptyonoprogne rupestris	breeding	N	5000	50000	NULL	p	different method	=	x	
A346	Pyrrhocorax pyrrhocorax	breeding	Y	100	300	NULL	p	no change	x	x	
A372	Pyrrhula pyrrhula	breeding	N	1000	8000	NULL	p	no change	x	x	
A118	Rallus aquaticus	breeding	N	50	250	NULL	p		u	x	
A132	Recurvirostra avosetta	breeding	Y	200	500	NULL	p	no change	x	x	
A318	Regulus ignicapilla	breeding	N	50000	100000	NULL	p	no change	u	x	
A249	Riparia riparia	breeding	N	10000	50000	NULL	NULL	no change	u	x	
A275	Saxicola rubetra	breeding	N	50	100	NULL	p	no change	x	x	
A276	Saxicola torquatus	breeding	N	500000	1000000	NULL	p	no change	-	x	
A361	Serinus serinus	breeding	N	1000000	5000000	NULL	p	no change	-	-	
A332	Sitta europaea	breeding	N	100000	500000	NULL	p	no change	+	x	
A857	Spatula clypeata	breeding	N	100	500	NULL	p	different method	u	x	
A193	Sterna hirundo	breeding	Y	10	20	NULL	p	no change	x	x	
A885	Sternula albifrons	breeding	Y	620	650	NULL	p	no change	x	x	

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species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend	
A209	Streptopelia decaocto	breeding	N	80000	5000000	NULL	**p**	different method	+	+	
A210	Streptopelia turtur	breeding	N	10000	25000	NULL	**p**	genuine change	-	-	
A219	Strix aluco	breeding	N	10000	20000	NULL	**p**	no change	u	x	
A352	Sturnus unicolor	breeding	N	500000	1000000	NULL	**p**	no change	=	x	
A311	Sylvia atricapilla	breeding	N	1000000	5000000	NULL	**p**	no change	+	+	
A310	Sylvia borin	breeding	N	100	500	NULL	**p**	no change	x	x	
A304	Sylvia cantillans - [excluding subalpina]	breeding	N	100000	500000	NULL	**p**	no change	-	x	
A309	Sylvia communis	breeding	N	50000	100000	NULL	**p**	no change	x	x	
A303	Sylvia conspicillata	breeding	N	2500	5000	NULL	**p**	different method	u	x	
A570	Sylvia hortensis s. str. - sensu stricto [excluding crassirostris]	breeding	N	5000	10000	NULL	**p**	no change	-	x	
A305	Sylvia melanocephala	breeding	N	1000000	5000000	NULL	**p**	no change	=	x	
A302	Sylvia undata	breeding	Y	100000	500000	NULL	**p**	no change	-	x	
A004	Tachybaptus ruficollis	breeding	N	1000	10000	NULL	**p**	different method	=	+	
A228	Tachymarptis melba	breeding	N	100	500	NULL	**p**		u	x	
A048	Tadorna tadorna	breeding	N	100	500	NULL	**p**	genuine change	+	+	
A128	Tetrax tetrax	breeding	Y	5008	12836	8900	**males**	genuine change	-	-	
A162	Tringa totanus	breeding	N	10	30	NULL	**p**	no change	u	x	
A676	Troglodytes troglodytes all others	breeding	N	500000	2000000	NULL	**p**	different method	=	=	
A283	Turdus merula	breeding	N	100000	1000000	NULL	**p**	no change	-	x	
A285	Turdus philomelos	breeding	N	5000	10000	NULL	**p**	no change	u	+	
A287	Turdus viscivorus	breeding	N	10000	100000	NULL	**p**	no change	u	+	

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species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend	
A213	Tyto alba	breeding	N	5000	10000	NULL	**p**	no change	-	x	
A232	Upupa epops	breeding	N	100000	500000	NULL	**p**	no change	=	=	
A419	Uria aalge ibericus	breeding	Y	0	1	NULL	**i**	no change	-	-	
A142	Vanellus vanellus	breeding	N	1	10	NULL	**p**	different method	x	x	
A893	Zapornia pusilla	breeding	Y	NULL	NULL	1	**cmales**	no change	x	x	
A898	Accipiter nisus all others	passage	N	250	750	NULL	**i**		N/A	N/A	
A294	Acrocephalus paludicola	passage	Y	25	200	NULL	**i**		N/A	N/A	
A197	Chlidonias niger	passage	Y	50	250	NULL	**i**		N/A	N/A	
A014	Hydrobates pelagicus	passage	Y	500	2000	NULL	**i**		N/A	N/A	
A166	Tringa glareola	passage	Y	100	300	NULL	**i**		N/A	N/A	
A054	Anas acuta	wintering	N	10604	34952	23607	**i**		+	+	
A052	Anas crecca	wintering	N	11253	27000	22023	**i**		u	u	
A053	Anas platyrhynchos	wintering	N	7773	30106	16003	**i**		-	-	
A043	Anser anser	wintering	N	1622	5116	2692	**i**		-	u	
A773	Ardea alba	wintering	Y	90	129	116	**i**		+	+	
A169	Arenaria interpres	wintering	N	1067	2378	1636	**i**		u	u	
A222	Asio flammeus	wintering	Y	50	150	80	**i**	no change	=	=	
A059	Aythya ferina	wintering	N	438	2092	1086	**i**		u	=	
A061	Aythya fuligula	wintering	N	79	301	179	**i**		u	u	
A144	Calidris alba	wintering	N	2091	4464	3054	**i**		+	F	

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species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend	
A149	Calidris alpina - all non-breeding populations	wintering	Y	34610	57920	40251	i		=	=	
A143	Calidris canutus	wintering	N	41	862	486	i		u	F	
A147	Calidris ferruginea	wintering	N	2	235	83	i		u	F	
A148	Calidris maritima	wintering	N	40	100	59	i	different method	x	x	
A145	Calidris minuta	wintering	N	184	441	331	i		-	u	
A861	Calidris pugnax	wintering	Y	7	166	77	i		u	+	
A138	Charadrius alexandrinus	wintering	Y	779	2540	1379	i		-	u	
A137	Charadrius hiaticula	wintering	N	5324	9652	6751	i		+	+	
A081	Circus aeruginosus	wintering	Y	500	1250	777	i	different method	+	+	
A480	Cyanecula svecica	wintering	Y	200	750	302	i	different method	+	x	
A026	Egretta garzetta	wintering	Y	542	1555	1063	i		u	x	
A098	Falco columbarius	wintering	Y	50	150	59	i	different method	+	x	
A125	Fulica atra	wintering	N	9970	25601	16229	i		=	u	
A126	Fulica cristata	wintering	Y	5	20	NULL	i		x	x	
A153	Gallinago gallinago	wintering	N	NULL	NULL	9000	i		-	x	
A127	Grus grus	wintering	Y	6591	11972	9171	i	no change	+	+	
A130	Haematopus ostralegus	wintering	N	256	20121	1272	i		u	-	
A131	Himantopus himantopus	wintering	Y	837	1933	1339	i		u	u	
A894	Hydroprogne caspia	wintering	Y	5	30	NULL	i		x	x	
A176	Larus melanocephalus	wintering	Y	4000	11000	6180	i	different method	x	x	

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species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend	
A157	Limosa lapponica	wintering	Y	557	7635	2819	i		u	u	
A156	Limosa limosa - all non-breeding populations	wintering	N	9963	32298	17759	i		=	u	
A152	Lymnocryptes minimus	wintering	N	NULL	NULL	2500	i		x	x	
A855	Mareca penelope	wintering	N	4110	14877	8806	i		=	u	
A889	Mareca strepera	wintering	N	876	3557	2334	i		u	u	
A900	Melanitta nigra s. str. - sensu stricto [excluding americana]	wintering	N	7600	13700	NULL	i	different method	x	x	
A069	Mergus serrator	wintering	N	1	68	33	i		u	-	
A074	Milvus milvus	wintering	Y	2000	2500	NULL	i	different method	-	x	
A016	Morus bassanus	wintering	N	180000	200000	NULL	i	different method	x	x	
A058	Netta rufina	wintering	N	148	492	335	i		u	=	
A768	Numenius arquata arquata	wintering	N	422	2512	1127	i		-	-	
A094	Pandion haliaetus	wintering	Y	150	200	NULL	i	different method	+	+	
A391	Phalacrocorax carbo sinensis	wintering	N	3641	7687	6447	i		u	u	
A663	Phoenicopterus roseus	wintering	Y	2875	8857	5625	i		u	+	
A034	Platalea leucorodia	wintering	Y	1363	2236	1798	i		+	+	
A140	Pluvialis apricaria	wintering	Y	NULL	NULL	NULL	NULL		+	x	
A141	Pluvialis squatarola	wintering	N	2331	9844	6254	i		-	-	
A008	Podiceps nigricollis	wintering	N	68	1378	382	i		u	+	
A693	Puffinus mauretanicus	wintering	Y	17000	20000	NULL	i	different method	x	x	

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species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend	
A132	Recurvirostra avosetta	wintering	Y	7515	14251	9911	i		=	=	
A155	Scolopax rusticola	wintering	N	NULL	NULL	17000	i		x	x	
A857	Spatula clypeata	wintering	N	13273	30677	20406	i		+	u	
A048	Tadorna tadorna	wintering	N	517	2603	1041	i		+	+	
A863	Thalasseus sandvicensis	wintering	Y	24	2013	126	i		u	x	
A162	Tringa totanus	wintering	N	1648	5166	3661	i		-	-	
A286	Turdus iliacus	wintering	N	NULL	NULL	NULL	NULL		N/A	N/A	
A284	Turdus pilaris	wintering	N	NULL	NULL	NULL	NULL		N/A	N/A	
A142	Vanellus vanellus	wintering	N	NULL	NULL	NULL	NULL		+	x	

Newly arrived species (ARR) and species extinct after 1980 (EXBA). These species are included in the statistics of the National Summary.

species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend
A079	Aegypius monachus	breeding	Y	29	31	NULL	p	genuine change	+	N/A
A094	Pandion haliaetus	breeding	Y	5	5	5	p	genuine change	+	N/A
A391	Phalacrocorax carbo sinensis	breeding	N	200	500	NULL	p		+	N/A
A032	Plegadis falcinellus	breeding	Y	500	2000	NULL	p		+	N/A

Bird species for which optional reports were provided (OPT & introduced). These are not included in the statistics of the National Summary.

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species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend
A765-X	Acridotheres cristatellus	breeding	N	100	1100	NULL	**p**	no change	u	x

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PORTUGAL – AZORES (PTAC)

Regularly occurring bird species (PRE). These are included in the statistics of the National Summary

species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend
A221	Asio otus	breeding	N	149	244	NULL	**p**		x	x
A387	Bulweria bulwerii	breeding	Y	50	70	NULL	**p**	no change	=	=
A087	Buteo buteo	breeding	N	NULL	NULL	1890	**p**	genuine change	=	x
A851	Calonectris borealis	breeding	Y	NULL	NULL	188000	**p**	no change	x	x
A138	Charadrius alexandrinus	breeding	Y	NULL	NULL	25	**p**		x	x
A206-X	Columba livia - feral populations	breeding	N	155407	559937	345866	**p**	different method	=	x
A206	Columba livia - wild populations	breeding	N	NULL	NULL	NULL	**NULL**		x	x
A421	Columba palumbus azorica	breeding	Y	42168	136149	93241	**p**	different method	+	x
A113	Coturnix coturnix	breeding	N	15404	22829	18459	**cmales**	no change	-	x
A269	Erithacus rubecula	breeding	N	119298	364713	288681	**p**	different method	=	x
A657	Fringilla coelebs all others	breeding	N	1840123	2885885	2386660	**p**	different method	+	x
A153	Gallinago gallinago	breeding	N	NULL	NULL	228	**p**	different method	=	x
A123	Gallinula chloropus	breeding	N	NULL	NULL	30	**p**		x	x
A874	Hydrobates castro	breeding	Y	665	740	NULL	**p**	improved knowledge	=	x
A884	Hydrobates monteiroi	breeding	Y	361	391	NULL	**p**	improved knowledge	x	x
A604	Larus michahellis	breeding	N	2705	4249	NULL	**p**	no change	x	x
A261	Motacilla cinerea	breeding	N	443843	787381	637153	**p**	different method	+	x

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species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend
A882	Onychoprion fuscatus	breeding	N	NULL	NULL	1	**p**	no change	x	x
A880	Puffinus lherminieri - [including baroli]	breeding	Y	895	1741	NULL	**p**	no change	x	x
A013	Puffinus puffinus	breeding	N	115	240	NULL	**p**	no change	x	x
A453	Pyrrhula murina	breeding	Y	257	841	485	**p**	genuine change	=	+
A317	Regulus regulus	breeding	N	45517	307960	141325	**p**	different method	x	x
A155	Scolopax rusticola	breeding	N	3247	3704	3461	**cmales**	no change	=	x
A450	Serinus canaria	breeding	N	1162117	2331398	1843889	**p**	different method	+	x
A192	Sterna dougallii	breeding	Y	535	1068	NULL	**p**	no info	F	F
A193	Sterna hirundo	breeding	Y	2419	3411	NULL	**p**	no info	F	F
A209	Streptopelia decaocto	breeding	N	335	86913	NULL	**p**	genuine change	+	x
A351	Sturnus vulgaris	breeding	N	440519	967404	730997	**p**	different method	+	x
A311	Sylvia atricapilla	breeding	N	349284	819438	493190	**p**	different method	-	x
A283	Turdus merula	breeding	N	606445	820827	685599	**p**	different method	+	x
A054	Anas acuta	wintering	N	NULL	NULL	NULL	**NULL**		x	x
A052	Anas crecca	wintering	N	NULL	NULL	NULL	**NULL**		x	x
A053	Anas platyrhynchos	wintering	N	NULL	NULL	NULL	**NULL**		x	x
A169	Arenaria interpres	wintering	N	NULL	NULL	156	**i**		x	x
A061	Aythya fuligula	wintering	N	NULL	NULL	NULL	**NULL**		x	x
A144	Calidris alba	wintering	N	NULL	NULL	50	**i**		x	x
A026	Egretta garzetta	wintering	Y	NULL	NULL	NULL	**NULL**		x	x

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species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend
A125	Fulica atra	wintering	N	NULL	NULL	NULL	**NULL**		x	x
A003	Gavia immer	wintering	Y	NULL	NULL	NULL	**NULL**		x	x
A186	Larus hyperboreus	wintering	N	NULL	NULL	NULL	**NULL**		x	x
A855	Mareca penelope	wintering	N	NULL	NULL	NULL	**NULL**		x	x
A141	Pluvialis squatarola	wintering	N	NULL	NULL	28	**i**		x	x
A857	Spatula clypeata	wintering	N	NULL	NULL	NULL	**NULL**		x	x

Newly arrived species (ARR) and species extinct after 1980 (EXBA). These species are included in the statistics of the National Summary

No data provided.

Bird species for which optional reports were provided (OPT & introduced). These are not included in the statistics of the National Summary

species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend
A110-X	Alectoris rufa - non-native populations	breeding	N	NULL	NULL	20	**p**		x	x
A364-X	Carduelis carduelis - non-native populations	breeding	N	75369	210277	163286	**p**		x	x
A363-X	Chloris chloris - non-native populations	breeding	N	NULL	NULL	25	**p**		x	x

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species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend
A620-X	Passer domesticus s. str. - sensu stricto [excluding italiae]; non-native populations	breeding	N	1284664	4745017	3456628	**p**	different method	+	x

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PORTUGAL - MADEIRA (PTMA)

Regularly occurring bird species (PRE). These are included in the statistics of the National Summary.

species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend
A401	Accipiter nisus granti	breeding	Y	43	99	NULL	**p**	improved knowledge	-	=
A432	Anthus berthelotii	breeding	N	10000	50000	NULL	**p**	no change	-	=
A227	Apus pallidus	breeding	N	1000	5000	NULL	**p**	no change	x	x
A425	Apus unicolor	breeding	N	5000	10000	NULL	**p**	no change	x	=
A387	Bulweria bulwerii	breeding	Y	NULL	45000	1	**p**	improved knowledge	+	+
A087	Buteo buteo	breeding	N	NULL	500	1	**p**	improved knowledge	=	=
A851	Calonectris borealis	breeding	Y	32864	43664	NULL	**p**	no change	+	+
A364	Carduelis carduelis	breeding	N	1000	5000	NULL	**p**	no change	=	+
A138	Charadrius alexandrinus	breeding	Y	0	50	NULL	**p**	no change	x	x
A363	Chloris chloris	breeding	N	1000	5000	NULL	**p**	no change	-	+
A206-X	Columba livia - feral populations	breeding	N	10000	50000	NULL	**p**	no change	x	x
A206	Columba livia - wild populations	breeding	N	10000	50000	NULL	**p**	no change	x	x
A455	Columba trocaz	breeding	Y	10000	14000	NULL	**i**	improved knowledge	=	=
A113	Coturnix coturnix	breeding	N	250	500	1	**cmales**	no change	=	=
A269	Erithacus rubecula	breeding	N	10000	50000	NULL	**p**	no change	+	=
A096	Falco tinnunculus	breeding	N	1000	5000	NULL	**p**	no change	-	+
A657	Fringilla coelebs all others	breeding	N	50000	100000	NULL	**p**	no change	+	=

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species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend
A125	Fulica atra	breeding	N	0	50	NULL	**p**	no change	+	+
A123	Gallinula chloropus	breeding	N	50	100	NULL	**p**	no change	+	+
A874	Hydrobates castro	breeding	Y	5000	NULL	1	**p**	no change	x	x
A604	Larus michahellis	breeding	N	3732	NULL	1	**p**	no change	-	F
A476	Linaria cannabina	breeding	N	500	1000	NULL	**p**	no change	x	-
A261	Motacilla cinerea	breeding	N	1000	5000	NULL	**p**	no change	=	=
A355	Passer hispaniolensis	breeding	N	1000	5000	NULL	**p**	no change	=	-
A389	Pelagodroma marina	breeding	Y	77770	110565	NULL	**p**	no change	-	=
A357	Petronia petronia	breeding	N	1000	5000	NULL	**p**	no change	x	=
A872	Pterodroma deserta	breeding	Y	160	180	NULL	**p**	no change	=	=
A385	Pterodroma madeira	breeding	Y	65	80	NULL	**p**	improved knowledge	=	+
A880	Puffinus lherminieri - [including baroli]	breeding	Y	1512	3837	NULL	**p**	improved knowledge	=	=
A013	Puffinus puffinus	breeding	N	1250	5000	NULL	**p**	no change	-	x
A652	Regulus madeirensis	breeding	N	50000	100000	NULL	**p**	no change	+	+
A155	Scolopax rusticola	breeding	N	NULL	162	1	**cmales**	no change	-	-
A450	Serinus canaria	breeding	N	10000	50000	NULL	**p**	no change	=	=
A478	Spinus spinus	breeding	N	1000	5000	NULL	**p**	no change	+	+
A192	Sterna dougallii	breeding	Y	0	50	NULL	**p**	no change	x	x
A193	Sterna hirundo	breeding	Y	500	1000	NULL	**p**	no change	x	x
A209	Streptopelia decaocto	breeding	N	100	500	NULL	**p**	no change	+	+
A311	Sylvia atricapilla	breeding	N	50000	100000	NULL	**p**	no change	+	=

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species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend
A303	Sylvia conspicillata	breeding	N	1000	5000	NULL	**p**	no change	x	=
A283	Turdus merula	breeding	N	50000	100000	NULL	**p**	no change	+	=
A213	Tyto alba	breeding	N	500	1000	NULL	**p**	no change	F	=
A232	Upupa epops	breeding	N	500	1000	NULL	**p**	no change	=	=

Newly arrived species (ARR) and species extinct after 1980 (EXBA). These species are included in the statistics of the National Summary.

No data provided.

Bird species for which optional reports were provided (OPT & introduced). These are not included in the statistics of the National Summary.

species code	species name	season	Annex I	population size min	population size max	population size best value	population size unit	reason for change	short-term population trend	long-term population trend
A110-X	Alectoris rufa - non-native populations	breeding	N	1000	5000	NULL	**p**	no change	x	x

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